Cefepime-taniborbactam (FTB) epidemiological cutoff values (ECOFFs) against key Enterobacterales species and Pseudomonas aeruginosa

A. Belley¹, M. Hackel², M. Wise², D. Sahm², C.L. Chatwin¹, B. Miller¹, A.K. Stevenson¹, M. Edwards¹, D.M. Daigle¹, G. Moeck¹

P1468 **ESCMID Global 2025 April 11-15, 2025** Vienna, Austria

¹Venatorx Pharmaceuticals, Inc., Malvern PA (United States) ²IHMA, Inc., Schaumburg, IL (United States)



Background

- Taniborbactam is an investigational cyclic boronate βlactamase inhibitor that restores cefepime activity against cefepime-, carbapenem-, and multidrug-resistant isolates of Enterobacterales and P. aeruginosa producing serineand metallo-β-lactamases (Hamrick 2020)
- Susceptibility breakpoints for new agents are informed by epidemiological cutoff values (ECOFFs) for the wild-type MIC distribution, which represent the highest MIC for organisms that lack phenotypically detectable acquired resistance mechanisms (EUCAST SOP 10.2)
- One objective of determining ECOFFs is to avoid splitting the wild-type MIC distribution when setting susceptibility breakpoints, thereby limiting excessive categorical errors due to inherent technical variation associated with MIC determinations (Arendrup 2009; Kahlmeter 2022)
- We determined cefepime-taniborbactam ECOFFs for clinical isolates of *Enterobacter cloacae* complex, Escherichia coli, Klebsiella pneumoniae, Proteus mirabilis, and P. aeruginosa

Results

- Six to eight cefepime-taniborbactam MIC distributions per species were aggregated (red curves; Figure 1) from 18,279 isolates of Enterobacterales and 9,607 isolates of P. aeruginosa
- The modeled wild-type MIC distributions for each of the five species (green curves; Figure 1) were primarily symmetrical and were either centered around the modal MIC or contained the modal MIC with a substantial shoulder 1 to 2 doubling dilutions higher than the mode
- Cefepime-taniborbactam ECOFFs to encompass ≥97.5% to ≥99.9% of the modeled wild-type populations ranged from 0.125 to 0.25 mg/L for Enterobacterales species and from 16 to 32 mg/L for *P. aeruginosa* (Table 1)

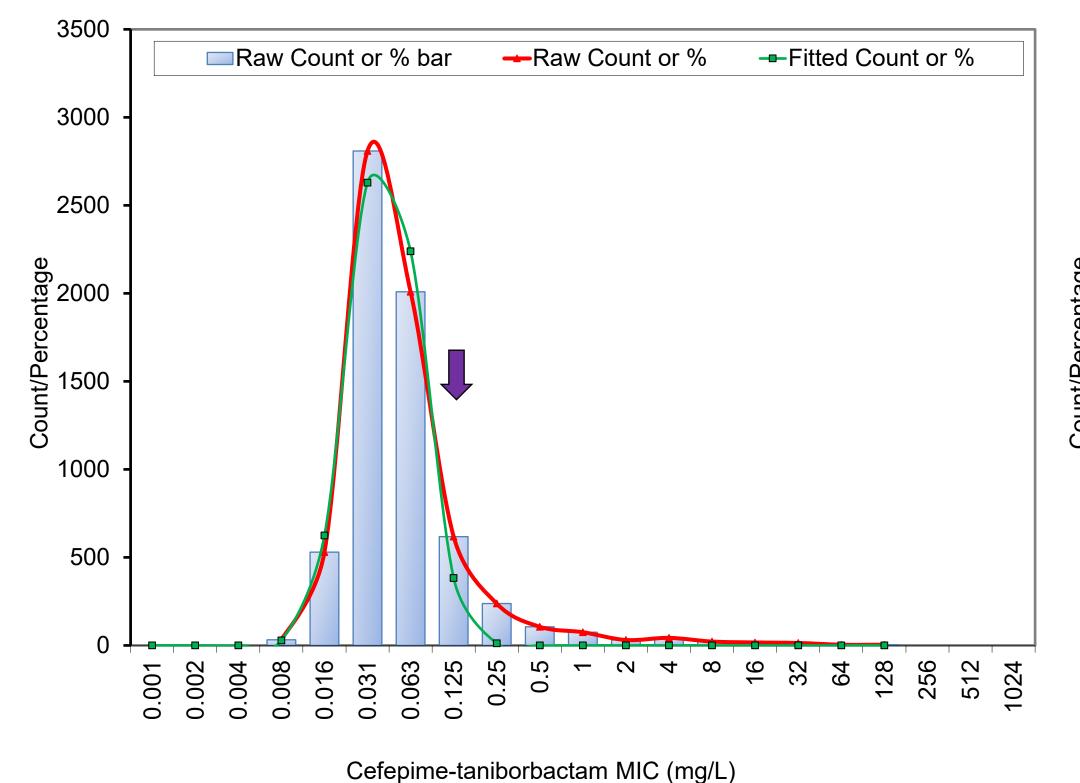
Conclusions

- A cefepime-taniborbactam ECOFF of 0.125 mg/L encompassed ≥99.0% of the modeled wild-type populations of *E. cloacae* complex, *E. coli*, *K.* pneumoniae, and P. mirabilis
- Whereas 92.6% of *P. aeruginosa* isolates in the aggregated MIC distribution were inhibited at ≤8 mg/L cefepime-taniborbactam, 17.3% of isolates had a cefepime-taniborbactam MIC = 8 mg/L
 - Given this 'shoulder' of isolates at 8 mg/L in the wildtype MIC distribution, a breakpoint of Susceptible ≤8 mg/L would split the wild-type distribution and lead to excessive categorical errors due to technical variation
 - Since only 3.5% of *P. aeruginosa* isolates in the aggregated distribution had a cefepime-taniborbactam MIC = 16 mg/L, a breakpoint of Susceptible ≤16 mg/L would largely alleviate this concern by leaving the wildtype MIC distribution intact
- A cefepime-taniborbactam ECOFF of 16 mg/L supports considerations for a breakpoint of Susceptible ≤16 mg/L for Enterobacterales species and *P. aeruginosa*, and is further supported by PK/PD analyses, mouse infection models (Abdelraouf 2020), and analyses of outcomes by MIC in the Phase 3 study CERTAIN-1 (Moeck 2024)

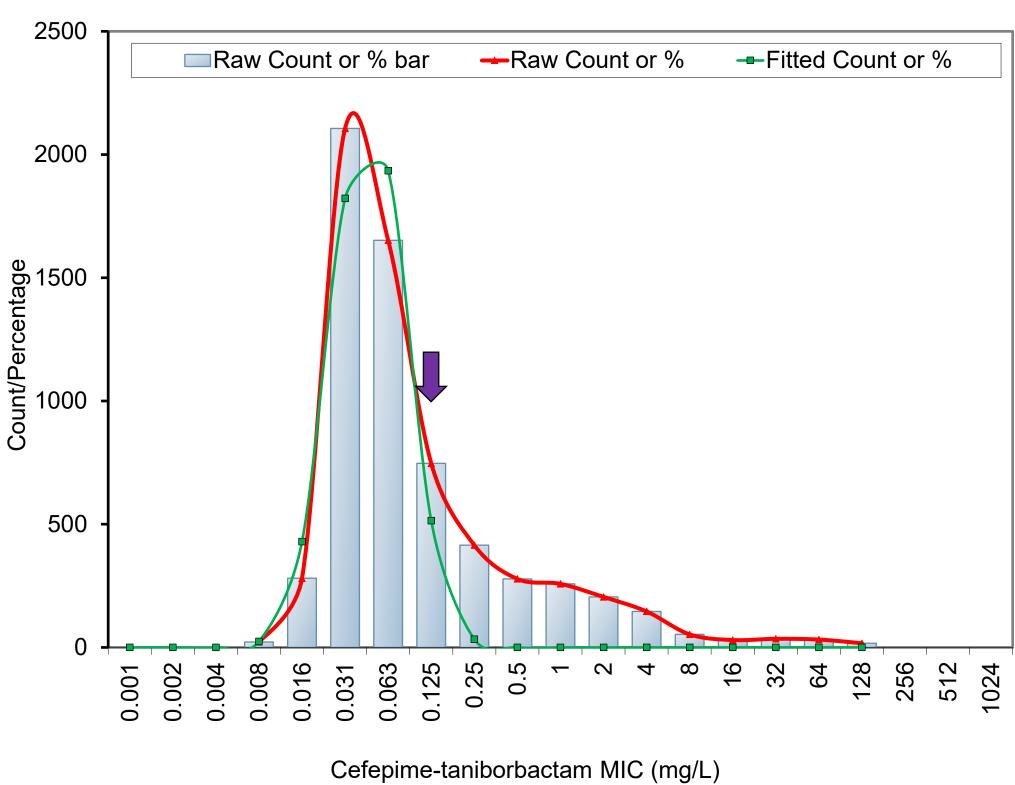
Results, continued

Figure 1. Statistical characterization of wild-type cefepime-taniborbactam MIC distributions for Enterobacterales **species and** *P. aeruginosa* (purple arrow in each distribution represents ECOFF value encompassing 99.0% of the wild-type population)

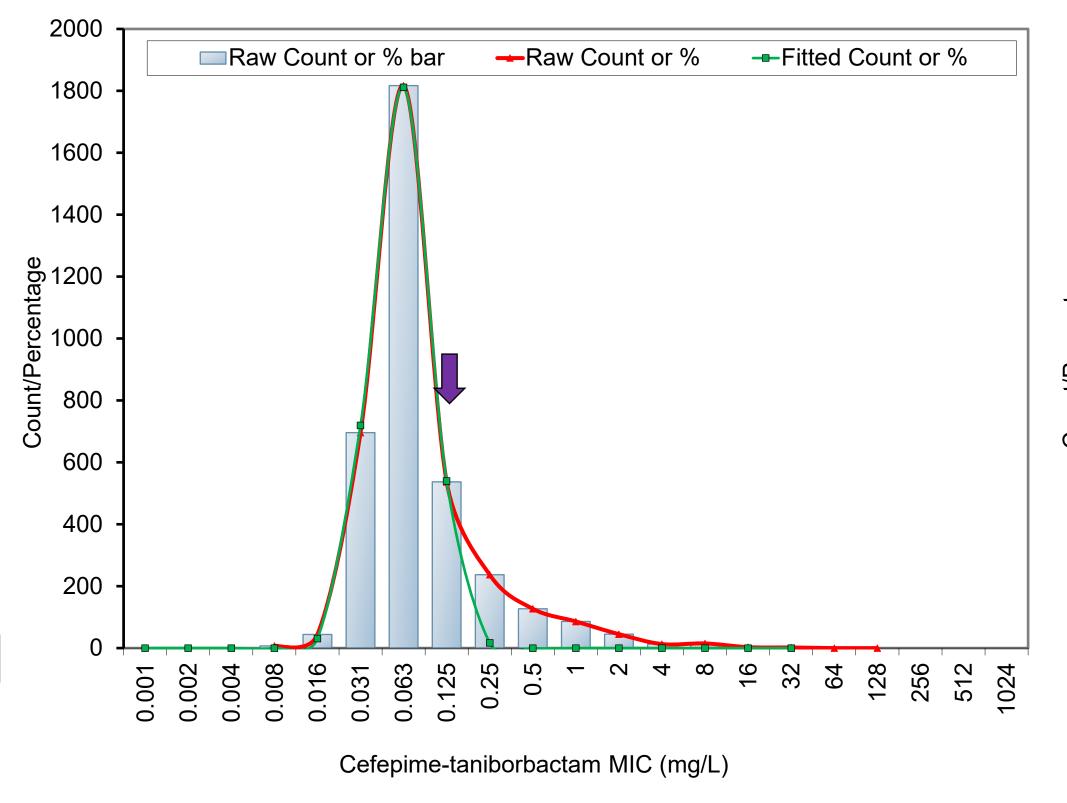
A. *E. coli* (n=6,552)



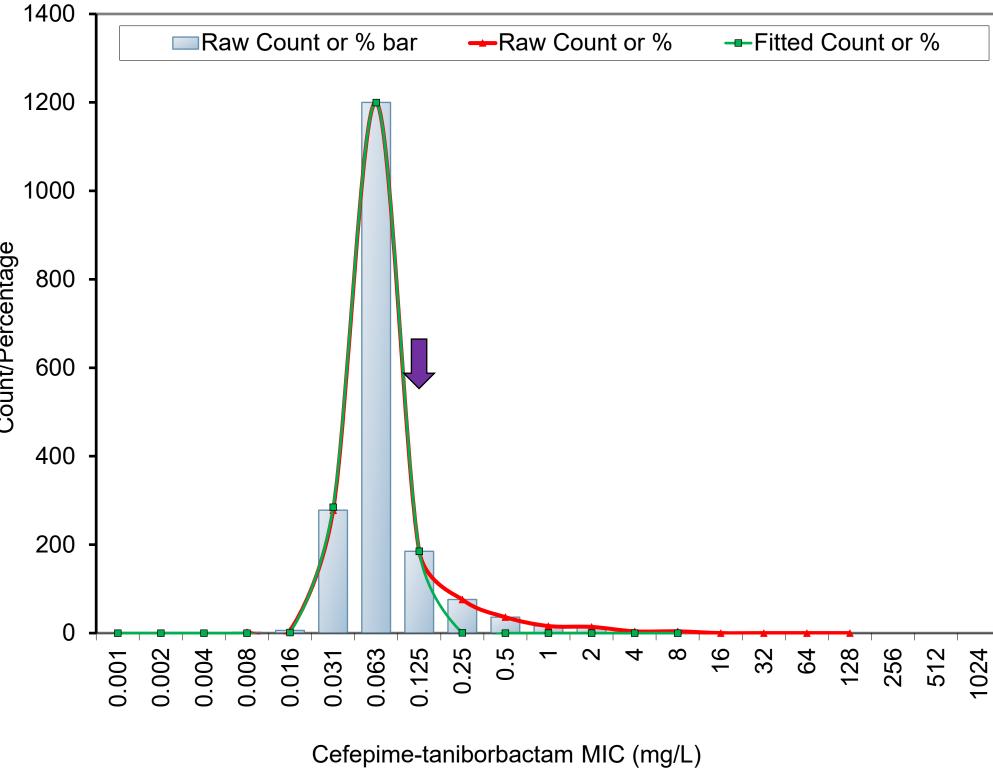
B. *K. pneumoniae* (n=6,277)



C. E. cloacae complex (n=3,629)



D. *P. mirabilis* (n=1,821)



E. *P. aeruginosa* (n=9,607)

-Fitted Count or % ■Raw Count or % bar --Raw Count or % 3000 2500 <u>g</u> 2000 날 1500 · 500 0.016 0.125 Cefepime-taniborbactam MIC (mg/L)

Purple arrow in each panel represents the ECOFF value encompassing 99.0% of the modeled wild-type population

Table 1. Cefepime-taniborbactam ECOFFs for *E.* cloacae complex, E. coli, K. pneumoniae, P. mirabilis, and *P. aeruginosa*, by %inhibition threshold

Organism (n)	Parameter	% inhibition threshold				
		95.0%	97.5%	99.0%	99.5%	99.9%
E. coli (6,552)	ECOFF (mg/L)	0.125	0.125	0.125	0.125	0.25
	%Obs > ECOFF	8.5%	8.5%	8.5%	8.5%	4.8%
	%Obs = ECOFF	6.5%	6.5%	6.5%	6.5%	0.2%
E. cloacae complex (3,629)	ECOFF (mg/L)	0.125	0.125	0.125	0.25	0.25
	%Obs > ECOFF	14.5%	14.5%	14.5%	8.0%	8.0%
	%Obs = ECOFF	17.3%	17.3%	17.3%	0.5%	0.5%
K. pneumoniae (6,277)	ECOFF (mg/L)	0.125	0.125	0.125	0.25	0.25
	%Obs > ECOFF	23.4%	23.4%	23.4%	16.8%	16.8%
	%Obs = ECOFF	10.8%	10.8%	10.8%	0.7%	0.7%
P. mirabilis (1,821)	ECOFF (mg/L)	0.125	0.125	0.125	0.125	0.125
	%Obs > ECOFF	8.2%	8.2%	8.2%	8.2%	8.2%
	%Obs = ECOFF	11.1%	11.1%	11.1%	11.1%	11.1%
P. aeruginosa (9,607)	ECOFF (mg/L)	8	16	16	32	32
	%Obs > ECOFF	7.4%	3.8%	3.8%	3.0%	3.0%
	%Obs = ECOFF	13.7%	3.6%	3.6%	0.5%	0.5%

ECOFF, epidemiological cutoff value (in mg/L)

%Obs > ECOFF, percentage of cefepime-taniborbactam MICs above the ECOFF %Obs = ECOFF, percentage of cefepime-taniborbactam MIC equal to the ECOFF.

References

- Abdelraouf K, Almarzoky Abuhussain S, Nicolau DP. In vivo pharmacodynamics of new-generation beta-lactamase inhibitor taniborbactam (formerly VNRX-5133) in combination with cefepime against serine-beta-lactamaseproducing Gram-negative bacteria. J Antimicrob Chemother. 2020 Dec 1;75(12):3601-3610. Arendrup MC, Kahlmeter G, Rodriguez-Tudela JL, Donnelly JP. Breakpoints for susceptibility testing should not divide wild-type distributions of important target species. Antimicrob Agents Chemother. 2009 Apr;53(4):1628-9
 - Clinical and Laboratory Standards Institute (CLSI). 2023. Development of In Vitro Susceptibility Test Methods, Breakpoints, and Quality Control Parameters, 6th Edition. Clinical and Laboratory Standards Institute (CLSI). 2018. Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria That Grow Aerobically, 11th Ed. CLSI Standard M07.
 - European Committee on Antimicrobial Susceptibility Testing (EUCAST) SOP 10.2. MIC distributions and the setting of epidemiological cutoff (ECOFF) values. December 2021. http://www.eucast.org/
- Hamrick JC, Docquier JD, Uehara T, Myers CL, Six DA, Chatwin CL, John KJ, Vernacchio SF, Cusick SM, Trout REL, Pozzi C, De Luca F, Benvenuti M, Mangani S, Liu B, Jackson RW, Moeck G, Xerri L, Burns CJ, Pevear DC, Daigle DM. 2020. VNRX-5133 (Taniborbactam), a Broad-Spectrum Inhibitor of Serine- and Metallo-β-Lactamases, Restores Activity of Cefepime in Enterobacterales and Pseudomonas aeruginosa. Antimicrob Agents Chemother 64(3):e01963-19. Kahlmeter G and Turnidge J. How to: ECOFFs—the why, the how, and the don'ts of EUCAST epidemiological cutoff values. Clin Microbiol Infect. 2022;28(7):952-954.

Karlowsky JA, Wise MG, Hackel MA, Six DA, Uehara T, Daigle DM, Pevear DC, Moeck G, Sahm DF. Cefepime-taniborbactam activity against antimicrobial-resistant clinical isolates of Enterobacterales and Pseudomonas

- aeruginosa: GEARS global surveillance programme 2018-22. J Antimicrob Chemother. 2024 Dec 2;79(12):3116-3131 Moeck G, Gasink LB, Mendes RE, Woosley LN, Dorr M, Chen H, Wagenlehner FM, Henkel T, McGovern PC. Patient outcomes by baseline pathogen resistance phenotype and genotype in CERTAIN-1, a Phase 3 study of cefepime-taniborbactam versus meropenem in adults with complicated urinary tract infection. Antimicrob Agents Chemother. 2024 Jul 9;68(7):e0023624.
- Wagenlehner FM, Gasink LB, McGovern PC, Moeck G, McLeroth P, Dorr M, Dane A, Henkel T; CERTAIN-1 Study Team. Cefepime-Taniborbactam in Complicated Urinary Tract Infection. N Engl J Med. 2024 Feb 15;390(7):611-

Methods

- Enterobacterales and *P. aeruginosa* isolates were from ongoing global surveillance between 2018 and 2023 (Karlowsky 2024), two additional independent surveillance studies, and a Phase 3 study in patients with complicated urinary tract infections (CERTAIN-1; Wagenlehner 2024).
- FTB MICs were determined by reference broth microdilution (ISO 20776-1:2019; CLSI 2018) at three central microbiology laboratories (IHMA, Schaumburg, IL; LabCorp, Indianapolis, IN; LabCorp, Shanghai, China) and at the Venatorx Pharmaceuticals laboratory (Malvern, PA).
- ECOFFs were determined for each species using ECOFFinder_XL_2010_v2.1 (https://www.eucast.org/fileadmin/src/media/PDFs/EUCAST files/MIC distributio ns/ECOFFinder XL 2010 v2.1 web version.xlsm) (EUCAST SOP 10.2 2021; CLSI 2023).